

## Effect of low temperatures on the infectivity of *Toxocara cati* larvae parasitized in mouse tissues

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### Summary

The infectivity of *Toxocara cati* larvae in mouse tissues, following storage at 4 °C and -25 °C, was measured using a bioassay in mice. Tissues of donor mice infected with 30-day-old *T. cati* larvae were stored at 4 °C for 7 and 14 days, and at -25 °C for 24 hours, then tissue larvae were inoculated into recipient mice (n = 6/group). After 15 days, larval burden in recipient mice was assessed by digestion. In the control group, a mean of 42.0 % of the inoculated larvae were established in mice. Storage of tissues at 4 °C for 7 and 14 days did not affect larval infectivity; means of 43.5 % and 37.4 % of inoculated larvae, respectively, were established in mice. Larvae exposed to -25 °C for 24 hours did not establish in mice, except for one larva recovered from a single mouse. These results may support the potential of tissue larvae in livestock animals as a causative agent of human toxocarosis.

Keywords: *Toxocara cati*; tissue larvae; cold tolerance; zoonosis

### Introduction

The common roundworm of cats, *Toxocara cati*, is an etiological agent of human toxocarosis with serious medical importance (Fisher, 2003; Akao & Ohta, 2007; Lee *et al.*, 2010). In addition to direct infection from eggs shed in the environment, cultural dietary preferences for raw or undercooked meat dishes may be associated with a risk of infection (Taira *et al.*, 2004). Accumulating evidence links human toxocarosis to the consumption of raw or undercooked meat (Mitsugi *et al.*, 1988; Nishikata *et al.*, 1991; Yoshikawa *et al.*, 2010; Okulewicz *et al.*, 2012), and an experimental study demonstrated that *T. cati* larvae persist and retain high infectivity in the muscles of chickens (Taira *et al.*, 2011).

As refrigeration or freezing of meat products are common

means of preservation throughout the consumer chain, an assessment of the risk associated with the consumption of *T. cati*-infected meat should include controlled experimental studies on the survival of larvae at low temperatures. It has been demonstrated that *T. cati* larvae in chicken muscle tissue retained infective even after 14 days of refrigeration at 4 °C, but also that deep freezing appeared to be an effective inactivation measure (Taira *et al.*, 2012).

Laboratory mice are easier to handle for experiments than chickens, particularly for studies to assess the infectivity of tissue larvae. In the present study, the infectivity of *T. cati* larvae in mouse tissues after storage at 4 °C and -25 °C was therefore investigated, and the results were compared with those obtained from previous study undertaken in the chicken model (Taira *et al.*, 2012).

### Material and methods

#### *Parasite isolates*

Eggs of *T. cati* were obtained from an experimentally infected cat. Fecal eggs were isolated, embryonated and stored in 1 % formalin at 25 °C for approx. 3 months (Taira *et al.*, 2012). The eggs were washed twice with tap water prior to inoculation to remove the formalin.

#### *Donor mice: infection and necropsy*

Twenty four 5-week-old male mice (outbred ICR strain) were inoculated orally with about 5500 embryonated *T. cati* eggs in 0.1 ml of an egg suspension. Food and water were administered *ad libitum*. All mice were euthanized and necropsied at 30 days post-infection.

Whole carcasses, excluding the skin, tips of limbs, tails, muzzles, stomach and intestines, were cut into pieces measuring less than approx. 5 mm<sup>3</sup> with a commercial food processor.

*Preservation of the mouse tissues*

The entire preparation of thoroughly mixed mouse tissue was divided into 12 portions of 50 g each, and these were placed in plastic bags. All portions of minced tissues in the plastic bags were flattened to about 2 cm in thickness. Three control bags were not stored, and the larval burden in these bags was assessed at the day of necropsy by HCl-pepsin digestion. Three sets of 3 bags were stored at 4 °C for 7 days, at 4 °C for 14 days, and at -25 °C for 24 hours, respectively. After storage, larvae in tissues were immediately released by digestion as described below.

*Recipient mice: infection and necropsy*

Twenty four 5-week-old male mice (outbred ICR strain) were used as recipient mice. Four groups of 6 mice were

each inoculated with around 100 larvae per mouse in approx. 0.5 ml saline, *per os* by stomach tubing. Inoculation was conducted within 1 hour after digestion to release the larvae. Fifteen days after inoculation, mice were euthanized. The whole carcass of each mouse, excluding the skin, tip of limbs, tail, muzzle, stomach, and intestines, was minced into pieces measuring approx. 5 mm<sup>3</sup> with a commercial food processor, and digested for larval recovery. The food processor was disinfected and washed with boiling water between the processing of each carcass.

All animals used in the study were treated in accordance with the guidelines for animal experimentation of Azabu University, and the relevant ethical guidelines of the Japanese Ministry of Education, Culture, Sports, Science and Technology.

Table 1. Infectivity of *Toxocara cati* larvae from mouse tissue after storage at 4 °C or -25 °C

Donor mouse tissues <sup>a</sup>	Recipient mice					
	Preservation temperature and duration	ID	Mean number of larvae inoculated (± 95 % CI)	Necropsy days after inoculation	Number of larvae recovered	Recovery %
			A		B	B/Ax100
0 days		1		15	33	35.9 %
		2			28	30.4 %
		3	92		29	31.5 %
		4	(± 9.4)		88	95.7 %
		5			37	40.2 %
		6			17	18.5 %
		Mean			38.7	42.0 %
4 °C, 7 days		7		15	29	29.6 %
		8			51	52.0 %
		9	98		48	49.0 %
		10	(± 8.2)		32	32.7 %
		11			50	51.0 %
		12			46	46.9 %
		Mean			42.7	43.5 %
4 °C, 14 days		13		15	35	35.4 %
		14			41	41.4 %
		15	99		41	41.4 %
		16	(± 12.7)		44	44.4 %
		17			10	10.1 %
		18			51	51.5 %
		Mean			37.0	37.4 %
-25 °C, 1 day		19		15	0	0.0 %
		20			1	0.9 %
		21	115		0	0.0 %
		22	(± 13.4)		0	0.0 %
		23			0	0.0 %
		24			0	0.0 %
		Mean			0.2	0 %

<sup>a</sup> Whole carcass, excluding the skin, tips of limbs, the tail, muzzle, stomach and intestines, of 24 mice inoculated with about 5500 embryonated *Toxocara cati* eggs, and necropsied 30 days after inoculation. The tissues were cut into pieces measuring less than approx. 5 mm<sup>3</sup>, and 50 g patties of tissue, each about 2 cm-thick, were preserved at temperatures and for durations, as described.

### Digestion and larval counts

The muscle digestion was conducted using the method of Taira *et al.* (2012). Briefly, the carcass of each mouse was digested in an HCl-pepsin solution for 2 hours. The ratio between tissue (g) and fluid (ml) was approximately 1:10. The digestion solution was sedimented for 1 hour and washed three times in 40 °C saline. Immediately after the final sedimentation, the recovered larvae were counted under a stereomicroscope.

### Results

No clinical signs of infection or changes in behavior were observed in any mice during the experiment. The infectivity of *T. cati* larvae in mouse tissues stored at 4 °C or -25 °C are presented in Table 1. A mean of 42.0 % ( $\pm$  27.26 SD) of the larvae obtained from donor mouse tissues at necropsy (controls) were recovered from the recipient mice. When administered tissues had been previously stored at 4 °C for 7 days, a mean of 43.5 % ( $\pm$  9.82 SD) of the inoculated larvae were established in recipient mice after 15 days. When tissues were stored at 4 °C for 14 days prior to inoculation of recipient mice, a mean of 37.4 % ( $\pm$  14.35 SD) of the inoculated larvae were recovered from recipient mice after 15 days. There were no statistically significant differences in recovery percentages of larvae among 0, 7 and 14 days of storages at 4 °C ( $p > 0.05$ , a one-way ANOVA with two-tailed test).

When the tissues containing larvae were stored at -25 °C for 24 hours, no larvae were recovered from 5 out of 6 recipient mice, with one larva being recovered from the remaining mouse at necropsy after 15 days.

### Discussion

The present study provides data on the infectivity of *T. cati* larvae in mouse tissue, after either refrigeration or deep freezing of the meat, to compare with those data from larvae in chicken meat (Taira *et al.*, 2012). The mouse bioassay employed in the present study substantiates that *T. cati* larvae retain high infectivity in mouse tissue stored at 4 °C for up to two weeks.

Taira *et al.* (2012) demonstrated that a mean of 47.1 % of larvae in fresh chicken meat were established in recipient mice, whereas a mean of only 24.1 % of the larvae present in chicken meat stored at 4 °C for 14 days were established. The authors assumed that this reduction in infectivity was a consequence of the decay of the meat, and the duration of exposure of the larvae to low temperature was not primarily impacted. The results of the present study support this assumption; the percent recovery of larvae in mouse tissue stored at 4 °C for 14 days (37.4 %) was not significantly reduced and was similar to that of the non-stored larvae used as control (42.0 %).

Deep freezing is regarded as an effective measure to inactivate *T. cati* larvae in mouse tissues, although one larva was herein recovered from a recipient mouse. Taira *et al.* (2012) reported that none of the larvae recovered after

freezing of chicken meat at -25 °C were able to establish in mice. The authors observed a few motile but non-infective larvae released from meat frozen for 12 hours. Given that one larva was found in a recipient mouse inoculated with tissue stored at -25 °C for 24 hours, further studies are required to determine the thermal dead point of *T. cati* larvae to derive the food safety implications.

In conclusion, the present report demonstrated that *T. cati* larvae in mouse tissues remained infective after 14 days of refrigeration at 4 °C, and that deep freezing dramatically reduced the larval infectivity. Given that survival of the larvae in mouse tissues stored at 4 °C was higher than survival of the larvae in chicken tissues subjected to the same treatment and tested in the same way (Taira *et al.*, 2012), screening of larvae in mouse tissue may be a useful model for an assessment of risk associated with the consumption of *T. cati*-infected meat.

### Acknowledgements

This study was partially supported by a research grant (Young Scientist Research Training Award 2012) funded by the Azabu University Research Service Division.

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RECEIVED FEBRUARY 5, 2013

ACCEPTED MARCH 15, 2013